

Challenges in Conducting Hazard Assessment of NP

BIO1: Environmental Fate and Possible Effects of
Nanoparticles

The 24th

Jyväskylä Summer School

Jyväskylän yliopisto



Nanoparticle Use



AT&T 3:41 PM

findNano

findNano

The Project on Emerging Nanotechnologies
at the Woodrow Wilson International Center for Scholars

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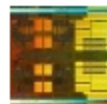
260 Den Nano Silver Far Infrared
Anti-odor Healthy Socks >



350TC Nano-Tex® Sheet Set by
Studio >



3XDRY® ESSEX SHIRT >



45nm Processors >



4Season OG Pants >



2009 Specialized Road Bike >

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All Products

Detail

APO-HG binocular

Minox

Date Added:
June 5, 2009

PRODUCT WEBSITE

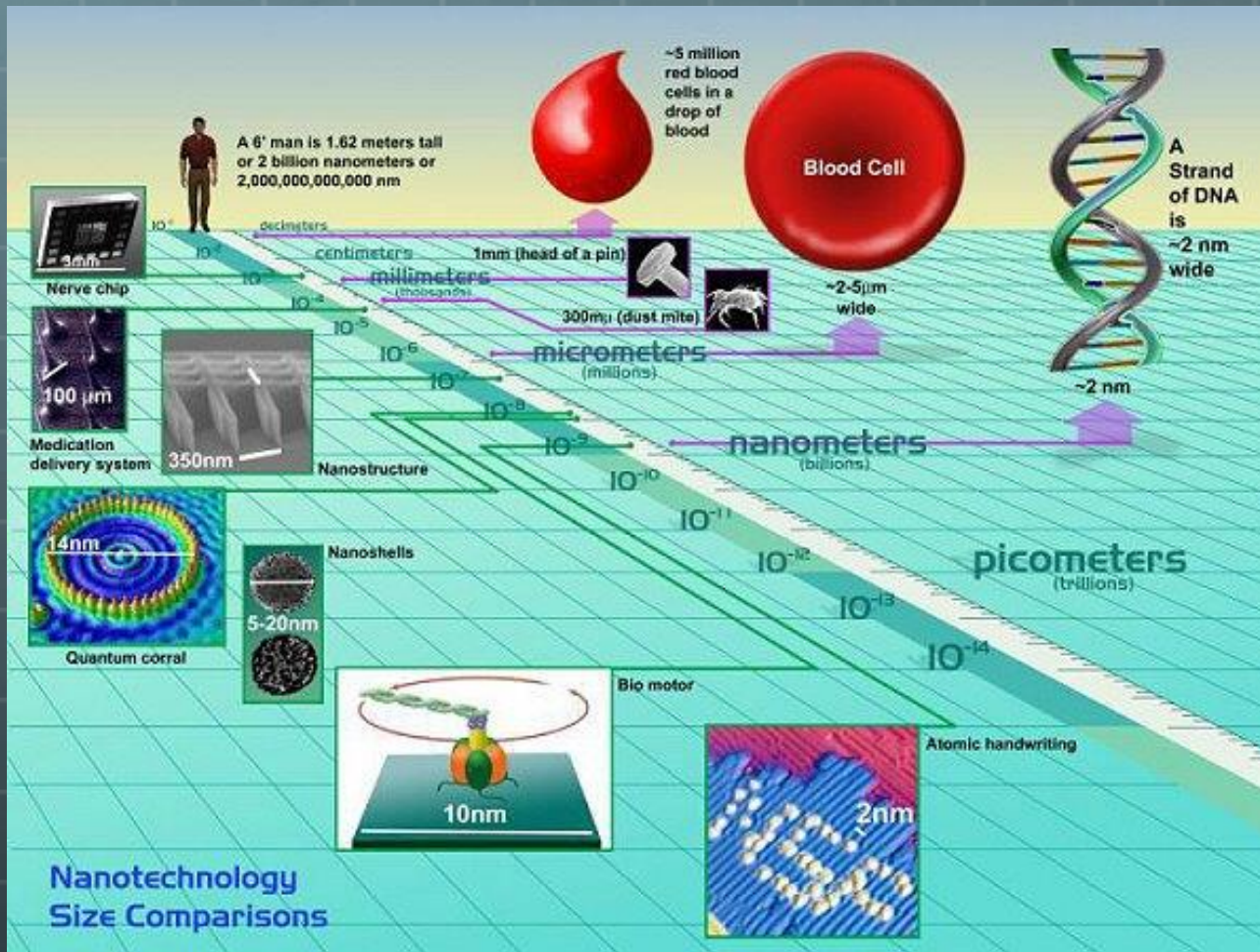


What They Say

SOURCE

"Pin-sharp optics with natural color rendition puts the new APO-HG binoculars from MINOX in a class of their own and first in the top league. The APO-HG 8.5x43 BR and the APO-HG 10x43 BR models impressively represent the latest technologies in binocular design. Featuring

Nanoparticles are NOT solutes



How do you measure concentration?

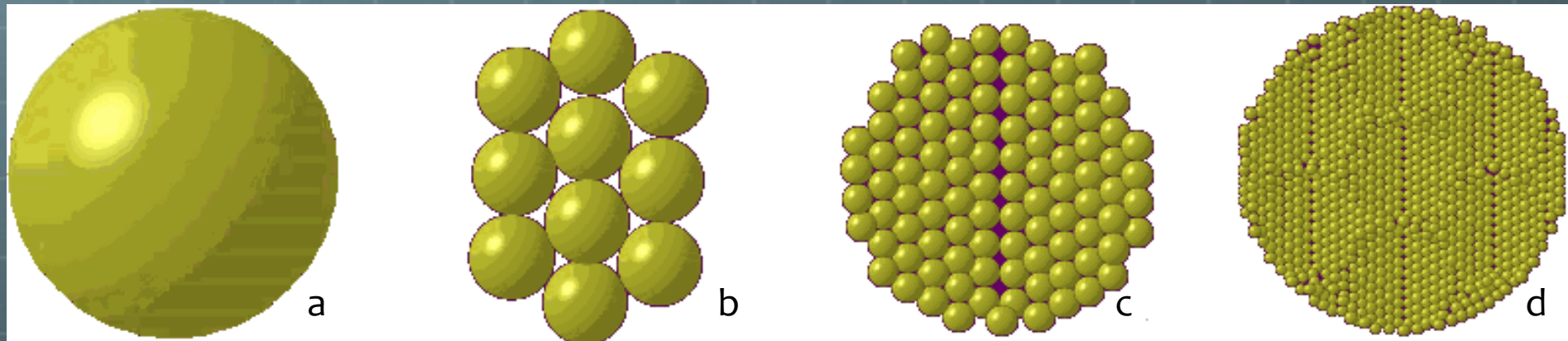
 mg/l

 Particles/l

 Surface area

 other





Particle Properties



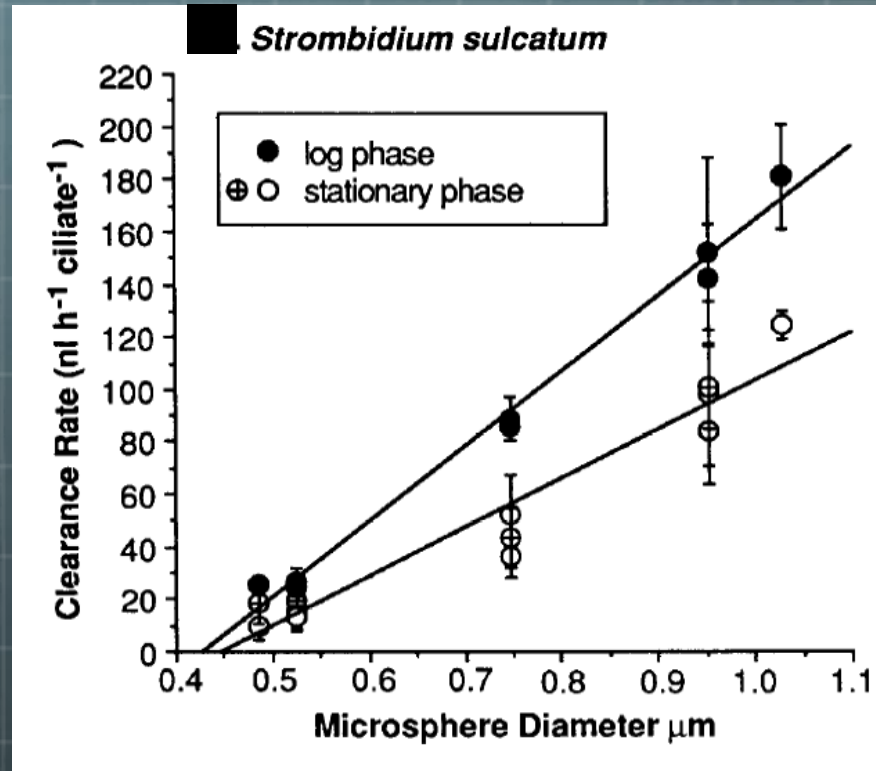
Gold Mass (μg)	1	1	1	1	1	1	1	1	1	1
Number of Particles	1 ^a	10 ^b	100 ^c	1,000 ^d	10,000	100,000	1,000,000	10,000,000	100,000,000	1,000,000,000
Particle Size (nm)	46,265.29	21,474.44	9,967.55	4,626.53	2,147.44	996.76	462.65	214.74	99.68	46.27
Particle Surface Area (nm ²)	6.72E9	1.45E10	3.12E10	6.72E10	1.45E11	3.12E11	6.72E11	1.45E12	3.12E12	6.72E12

Nanoparticle Behavior

Challenges

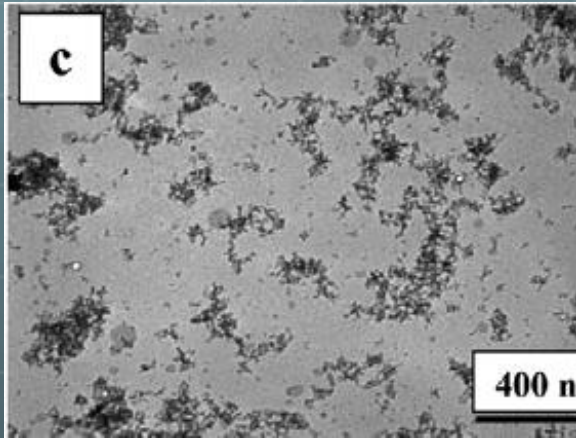
-  Batch to batch variability
-  Not truly soluble: colloidal suspension
-  Quantification
 -  Separating particles from ions and natural background concentrations (use of stable isotopes for Zn, Cu, different analysis techniques)

Nanoparticles in Aquatic Ecosystems

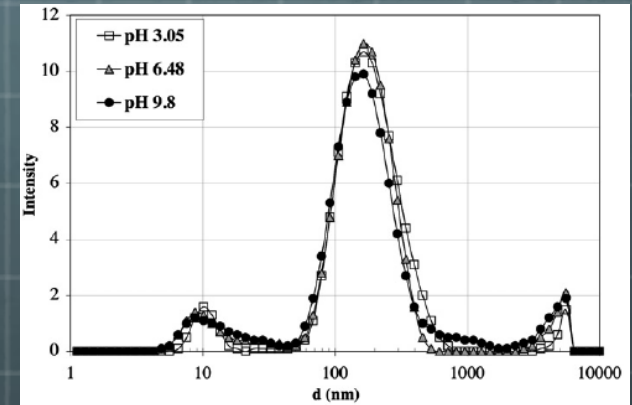


Nanoparticle size matters to filter-feeders
(Marine ciliate) Christaki et al 1998

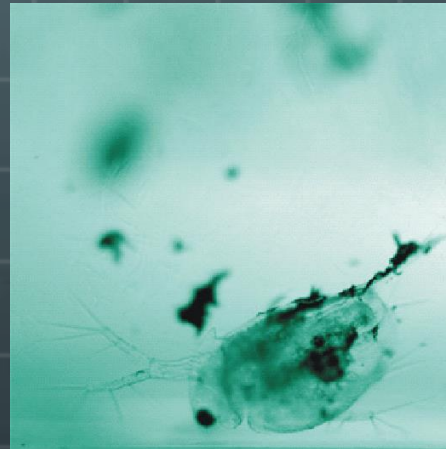
Nanoparticles in Aquatic Ecosystems



Particle Aggregation
(Fullerols) Brant et al 2007



Particle Size Distribution
(Fullerols) Brant et al 2007



Particle Stability
(SWNT) Roberts et al 2007

Nanoparticles in Aquatic Ecosystems

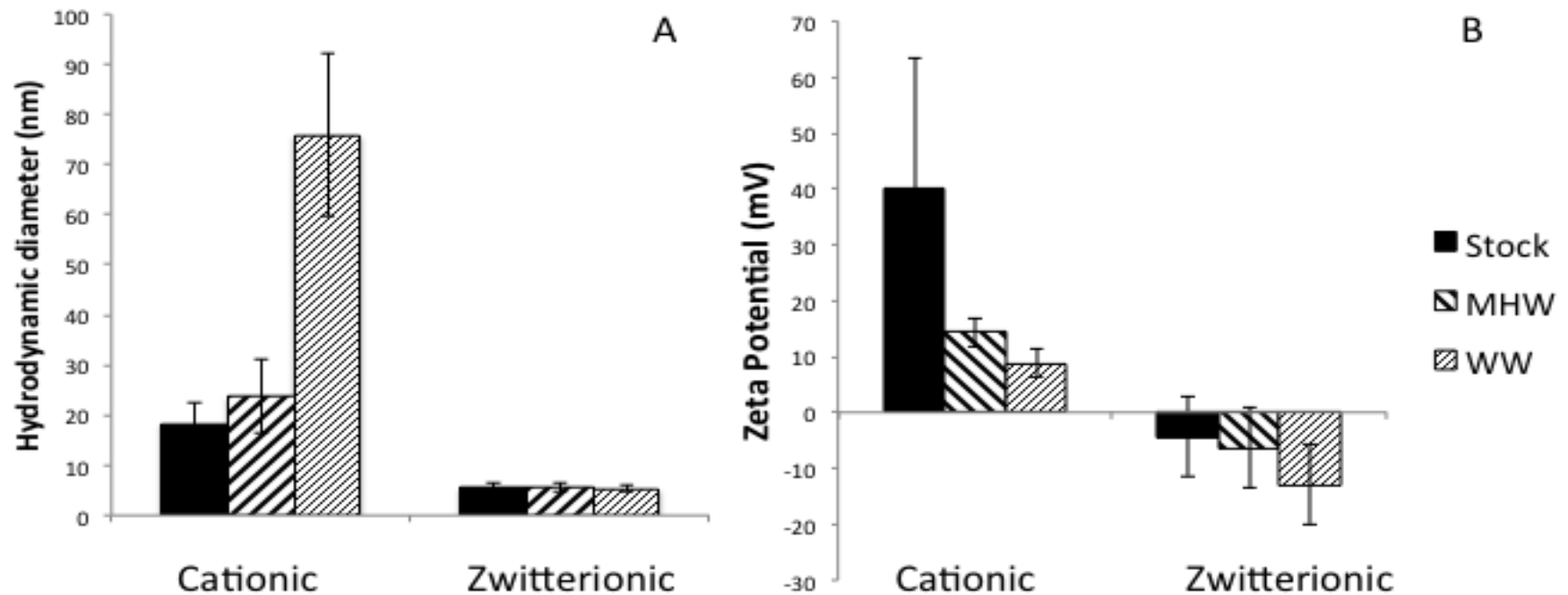


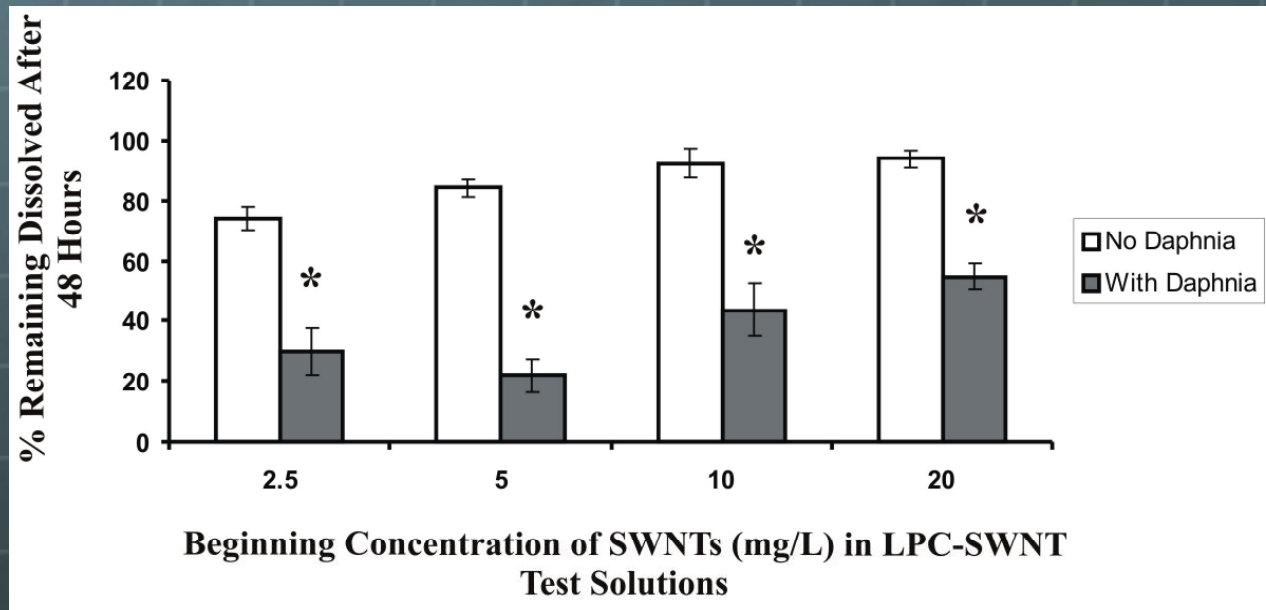
Figure 2. Hydrodynamic diameter (a) and zeta potential (b) for both nanoparticles in stock solution, moderately hard water and wastewater (after 1 hour). Each bar represents the average of three runs ± 1 standard deviation.

Aqueous conditions can also modify particle characteristics

Wray et al 2014, *In Prep*




Nanoparticles in Aquatic Ecosystems

Filter-feeders modify nanoparticle suspensions



(SWNT) Roberts et al 2007

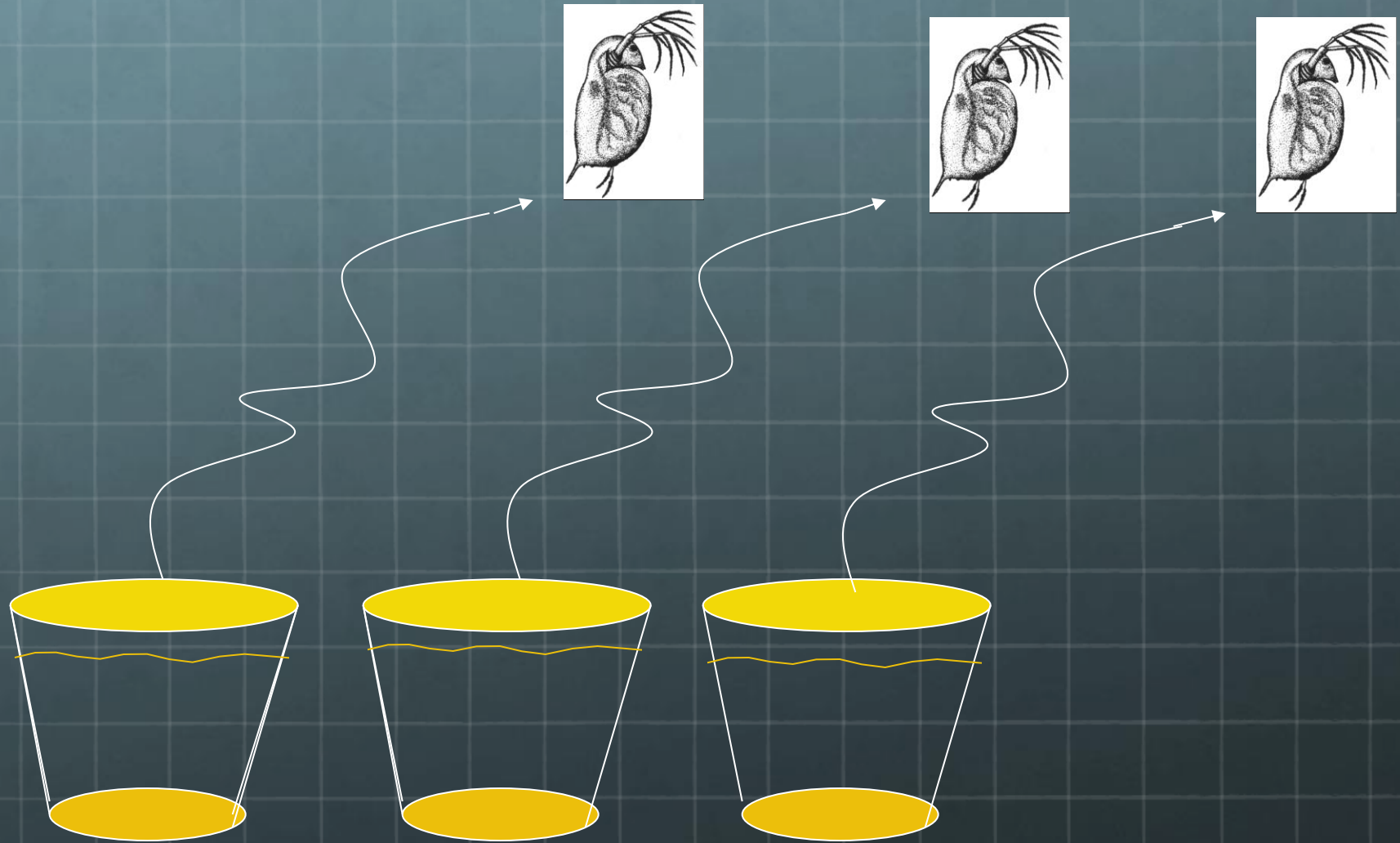
How do you test the effects of particles?

-  Static bioassays
-  Static renewal bioassays
-  Flow-through bioassays

Static Bioassay

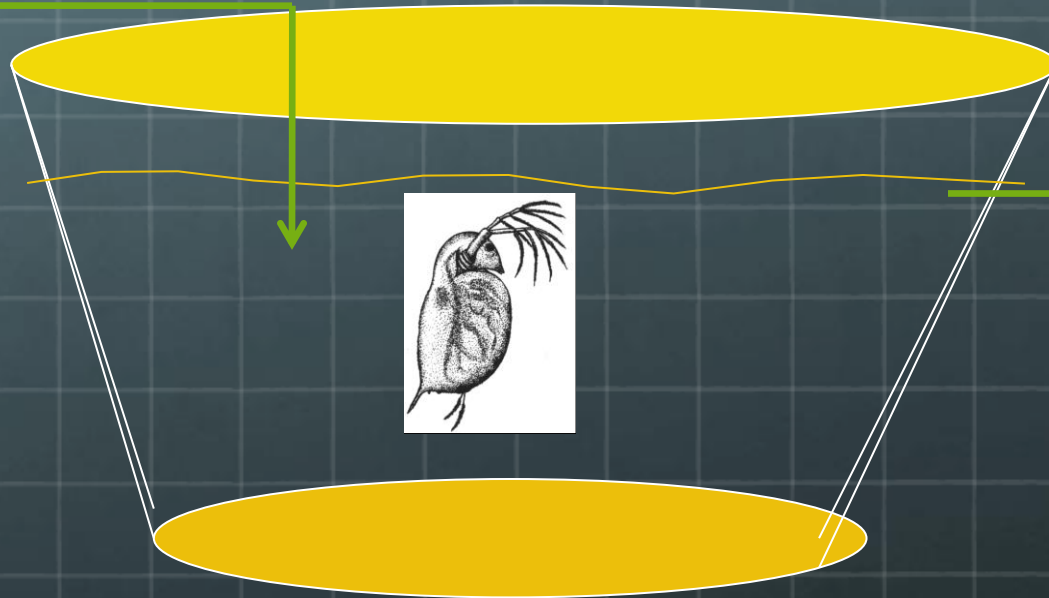


Static-Renewal Bioassay



Flow-through Bioassay

In Flow



Outflow

Current Environmental Research

- 🌐 Research publications on environmental fate and effects increasing exponentially
- 🌐 Initial research pointed out potential issues but also raised questions about research methods.
- 🌐 Not enough to simply report how much was put into the test medium.
 - 🌐 Size
 - 🌐 Shape
 - 🌐 Surface chemistry
 - 🌐 Aggregation rate

Experimental Design

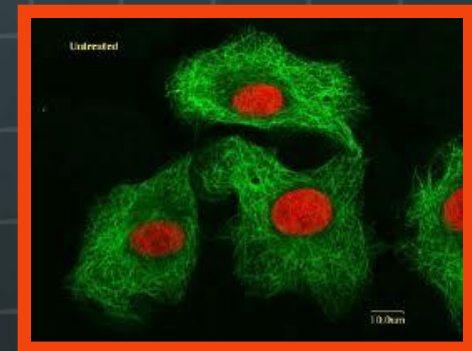
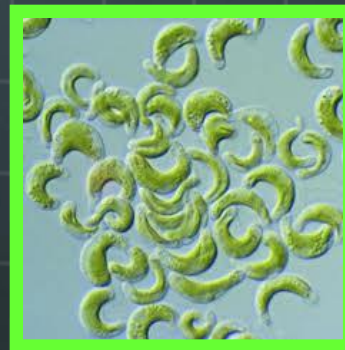
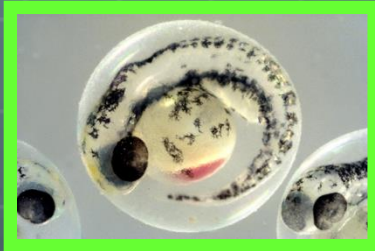
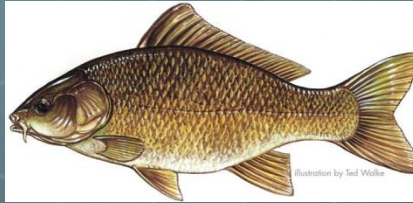
- Depends on the question
 - How toxic is the nanoparticle?
 - What is the mechanism of toxicity?
 - Can toxicity be attributed to the particle itself or is it a function of particle dissolution (particularly important for Ag, Cu and ZnO)?
 - What is the influence of size, shape, or surface chemistry on nanoparticle toxicity?
 - Does toxicity change based on the exposure media?
 - pH
 - Hardness
 - DOC
 - How does toxicity change between species?

How toxic?

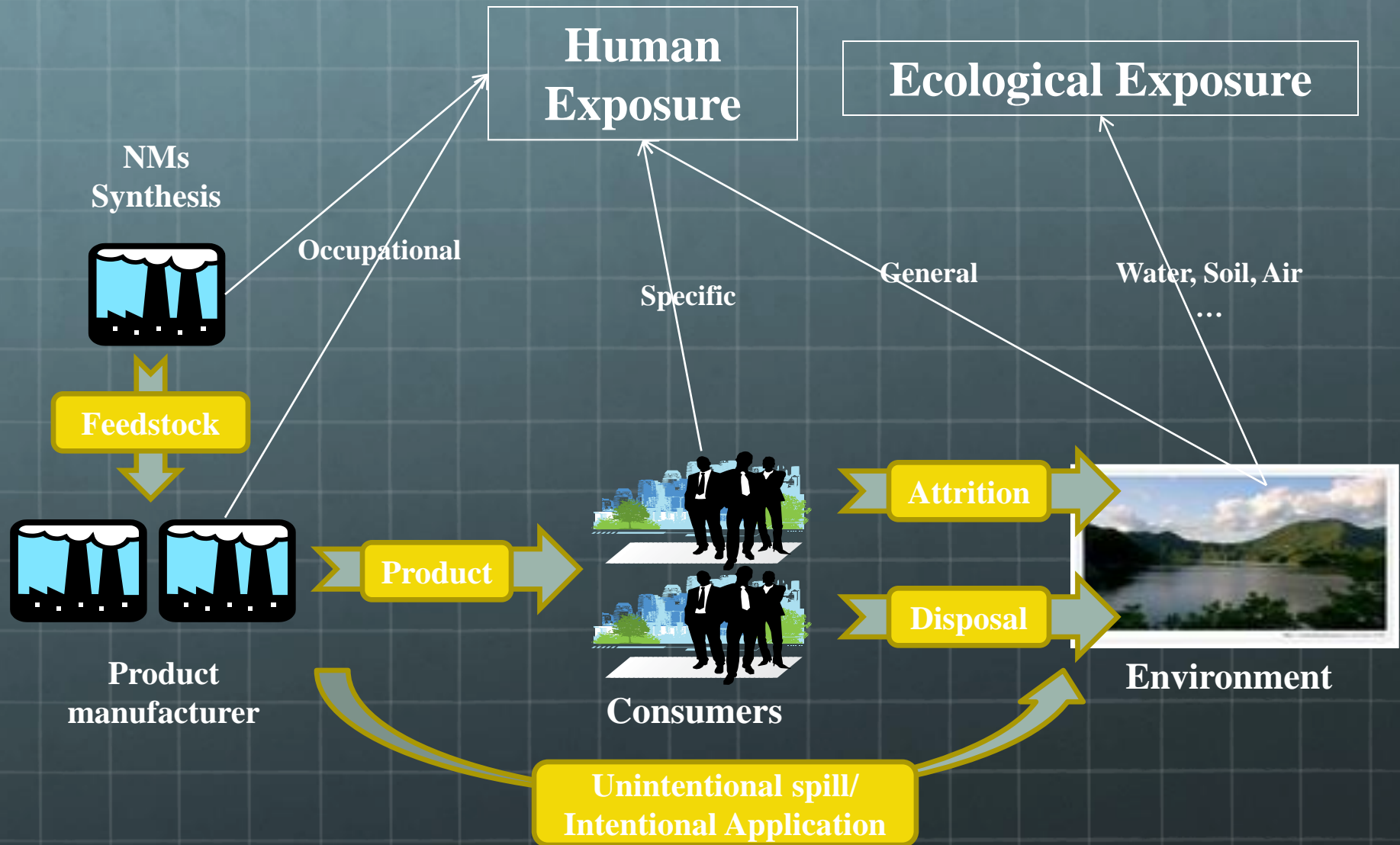
- Start with a standard bioassay procedure.
- Try to achieve constant exposure.
- Characterize particles during exposure
 - Do they aggregate, agglomerate (reversible), sediment, sorb to other surfaces, adsorb organic compounds, etc.

ENTOX Research

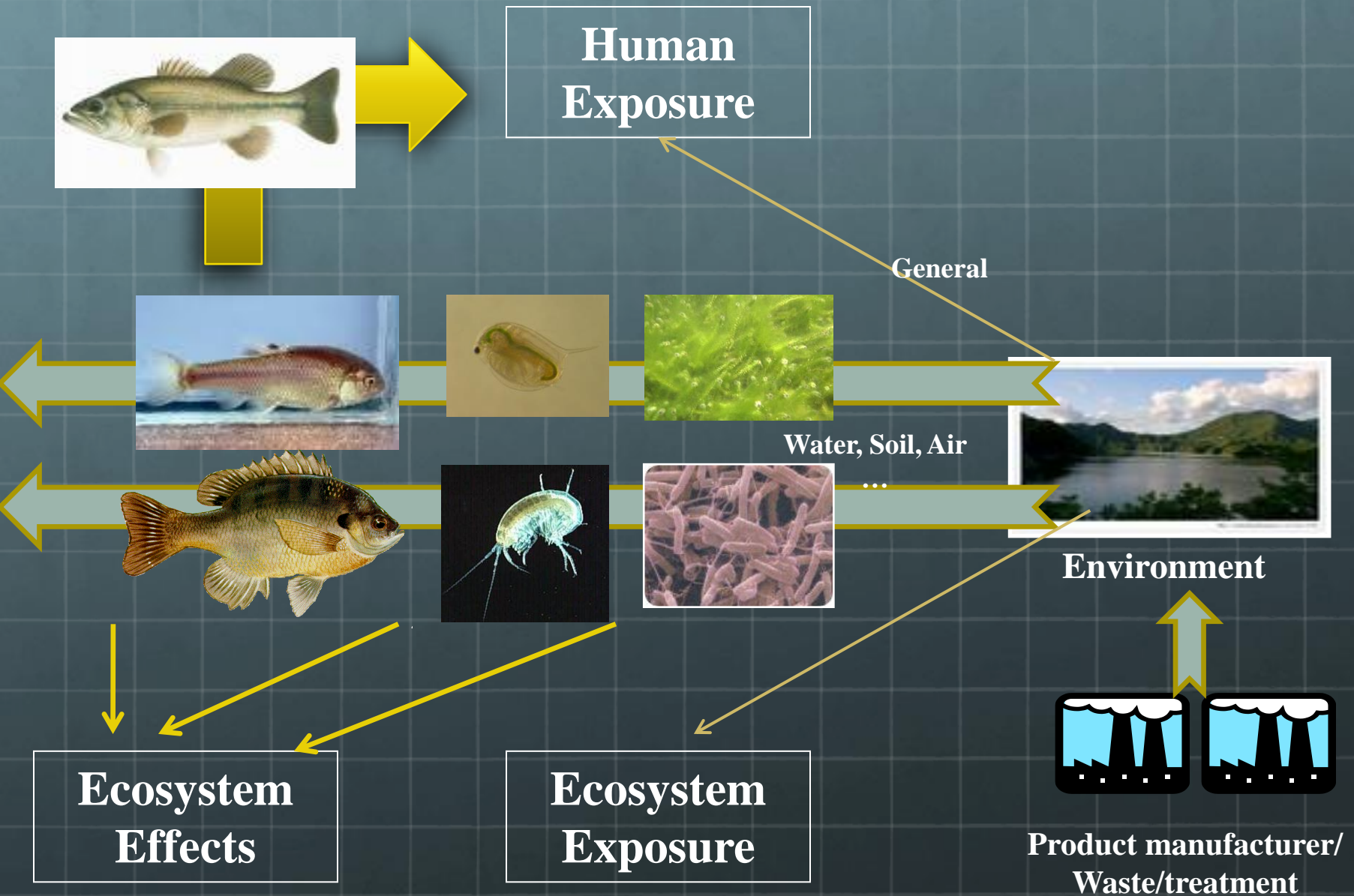
Complexity



Release and Exposure of Nanoparticles



Why Aquatic Toxicology?



What is the Influence of Nanoparticle Characteristics?

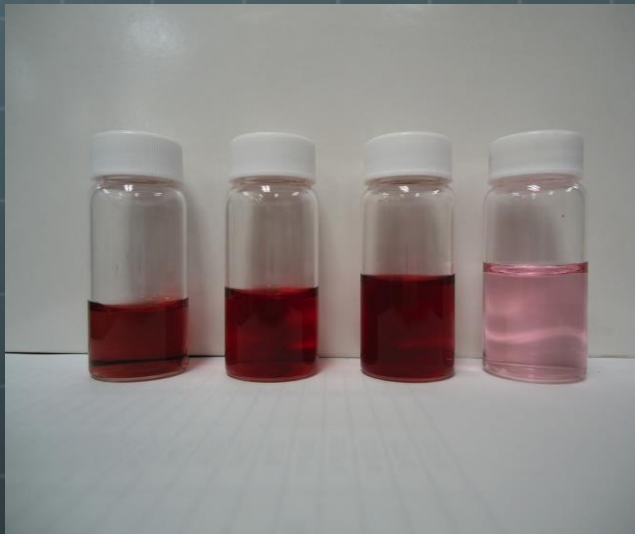
- Choose nanomaterial
- Choose characteristics
 - Run a factorial design
 - e.g. 3 sizes, 3 shapes, 3 surface chemistries
 - $3 \times 3 \times 3 = 27$ treatments
- Do characteristics change based on the exposure media?

What is the Influence of Particle Characteristics on the Toxicity of Gold Nanoparticles

Particle Core	Shape	Size	Surface Charge	# treatments/ Bioassay
Gold	Sphere	4, 10, 50 nm diameter	Amine (protonated) (cationic)	3
			Biotin (nonionic)	3
			Carboxylate (anionic)	3
	Cube	50, 75 nm	Amine (protonated) (cationic)	2
			Biotin (nonionic)	2
			Carboxylate (anionic)	2
	Rod	20 by 100 nm	Amine (protonated) (cationic)	2
		20 by 400 nm	Biotin (nonionic)	2
			Carboxylate (anionic)	2

Stock Solutions of In-House AuNPs

Shape	Sphere	Sphere	Sphere	Rod	Rod
Particle Size (nm)	5.67 ± 1.28	21.25 ± 2.5	30.64 ± 6.00	W: 17.82 ± 2.03 L: 58.08 ± 5.31	W: 17.82 ± 2.03 L: 58.08 ± 5.31
Surface Chemistry	Citrate	Citrate	Citrate	Poly(acrylic acid)	Poly(allylamine hydrochloride)
Zeta Potential (mV)	-39.8 ± 9.94	-35.7 ± 19.5	-38.9 ± 16.4	-20.7 ± 9.33	$+38.8 \pm 17.5$



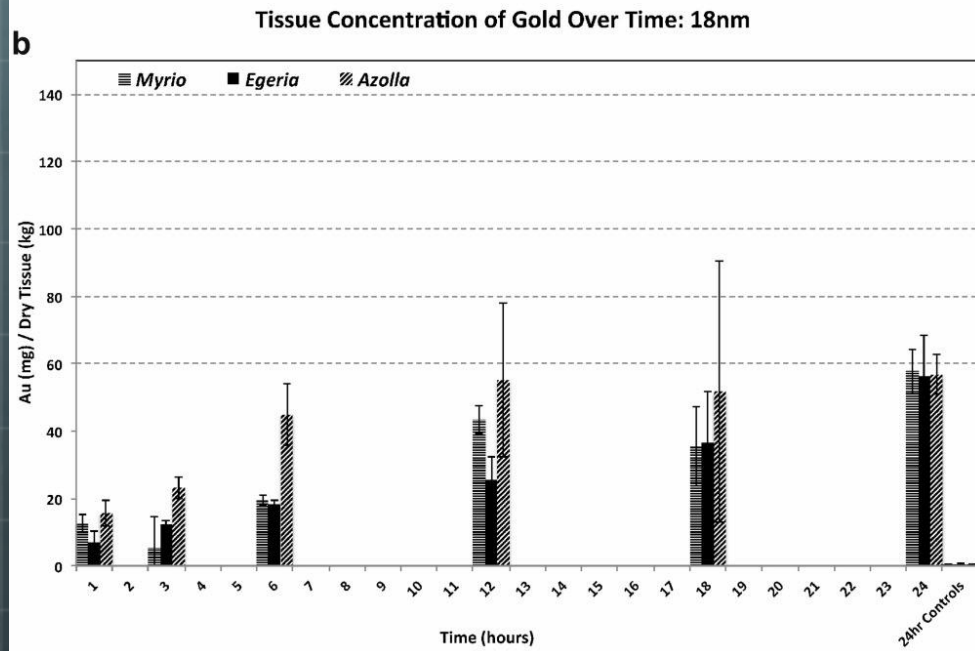
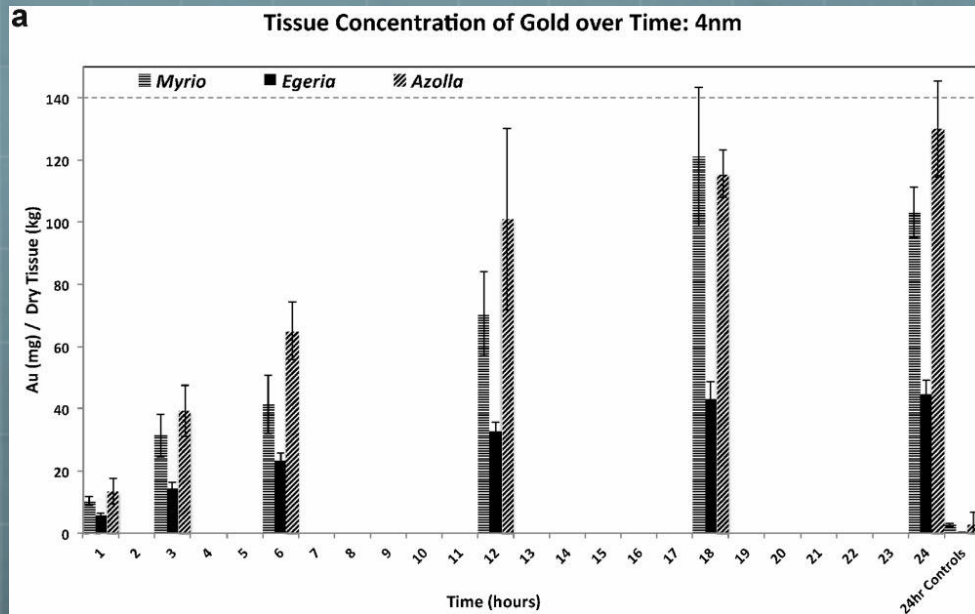
Size (nm)	Shape	Surface Ligand	$k_{uw} (\text{L g}_{\text{org}}^{-1} \text{d}^{-1})^a$	$k_e (\text{d}^{-1})$
4	Sphere	Citrate	5.139 ± 0.388	2.929 ± 0.140
20	Sphere	Citrate	2.772 ± 0.247	1.840 ± 0.190
30	Sphere	Citrate	2.679 ± 0.120	1.119 ± 0.213
18 x 58	Rod	Poly(acrylic acid)	1.548 ± 0.038	2.025 ± 0.287
18 x 58	Rod	Poly(allylamine hydrochloride)	L: 4.632 ± 0.830 H: 92.494 ± 6.504	2.746 ± 0.303

Individual rate constants for each particle configuration

Diameter (nm)	Surface Charge	$k_{uw} (\text{L g}_{\text{org}}^{-1} \text{d}^{-1})$	$k_e (\text{d}^{-1})$	BCF
4	Anionic	5.14	2.93	1750 ^a
20	Anionic	2.81	1.93	1460 ^a
30	Anionic	2.81	1.12	2510 ^a
4	Cationic	L: 5.14 H: 94.83	3.74	L: 1370 H: 25400
20	Cationic	L: 2.80 H: 92.02	2.75	L: 1020 H: 33500
30	Cationic	L: 2.80 H: 92.02	1.93	L: 1450 H: 47700

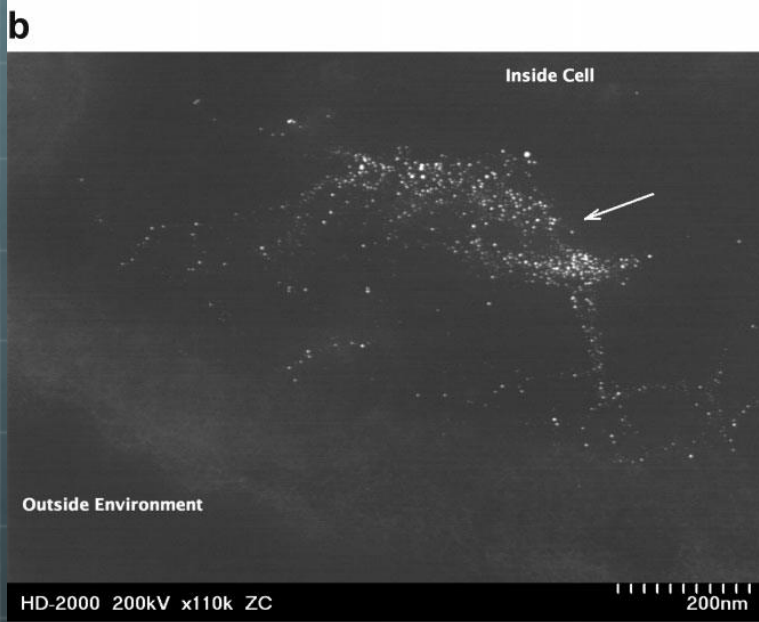
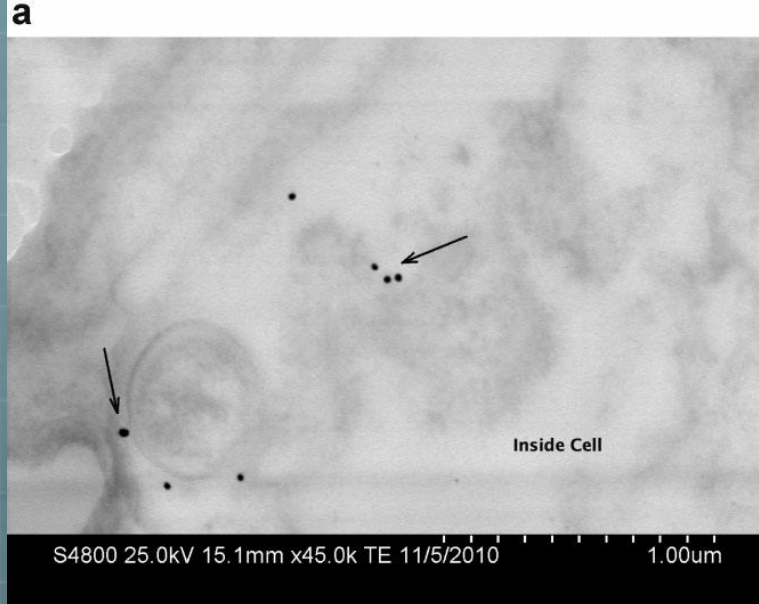
Model predictions for rate constants and BCF using full data set and multiple linear regression analysis

Species Differences

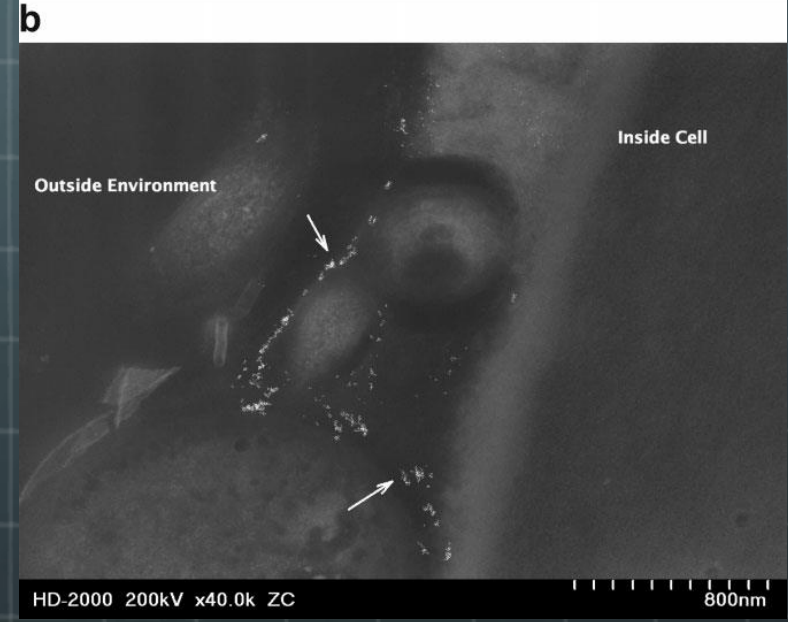
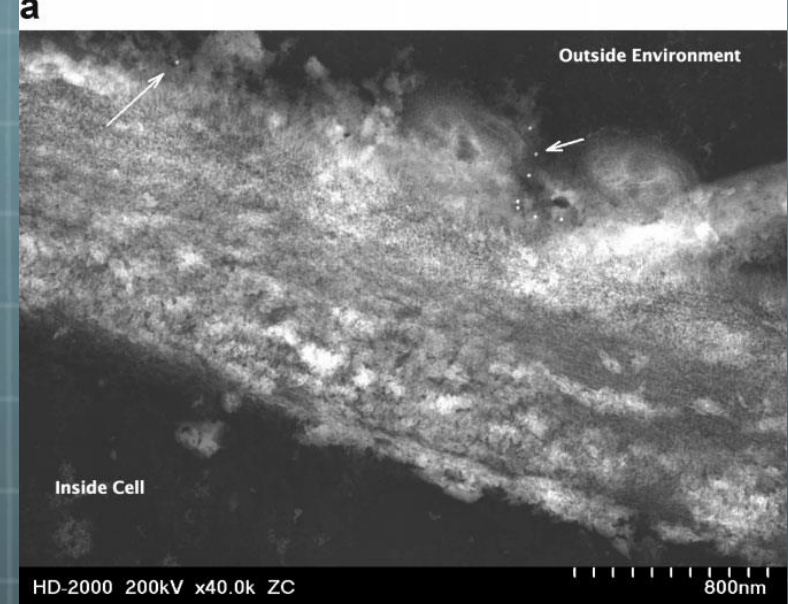


- Aquatic macrophytes exposed to citrate coated gold nanoparticles with 4 and 18 nm diameter

- Accumulation is both size and species dependent



A. caroliniana exposed to 18 (a) and 4 nm (b) gold NPs

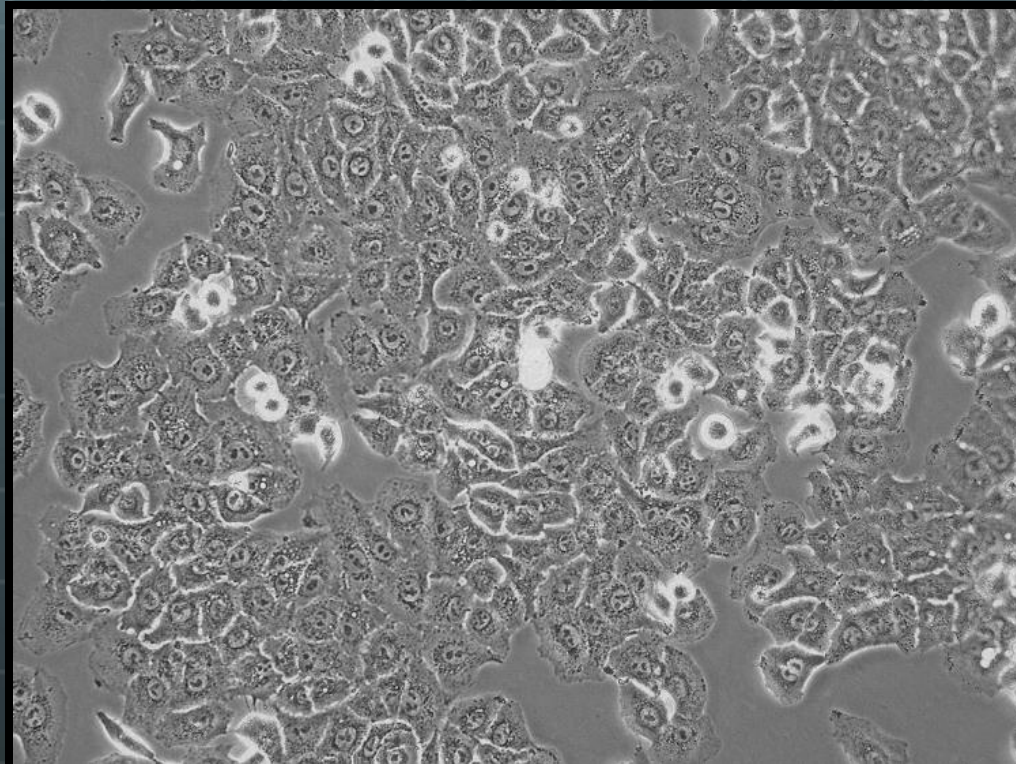


E. densa exposed to 18 (a) and 4 nm (b) gold NPs

Influence of Exposure Media

■ A549 Cells

- human carcinomic alveolar epithelial cells
- Squamous Cells extracted in 1972



Cell Culture Methods

- **Dulbecco's modified Eagle's Media**
 - 10 Fetal Bovine Serum (FBS)
 - 2mM Glutamine
 - 1% Non-essential AA's
 - 2% Penicillin/Streptomycin
- **75cm² TC-Treated Flask**
- **5% CO₂ at 37 °C**



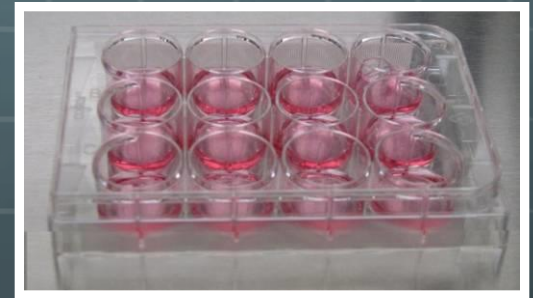
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Experimental Methods

- 12-well TC-Treated Plates
 - 10^5 cells per well
- 24hr for Attachment
- Fresh Media + Particles
- $n=3$
- Rinsed with PBS (x3)
- 40% aqua regia
- Analyzed ICP-MS



Primary Objective

To develop a rapid, high volume bioassay to facilitate investigations of a large array of particle modifications

1. Exposure Media

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1. Exposure Media

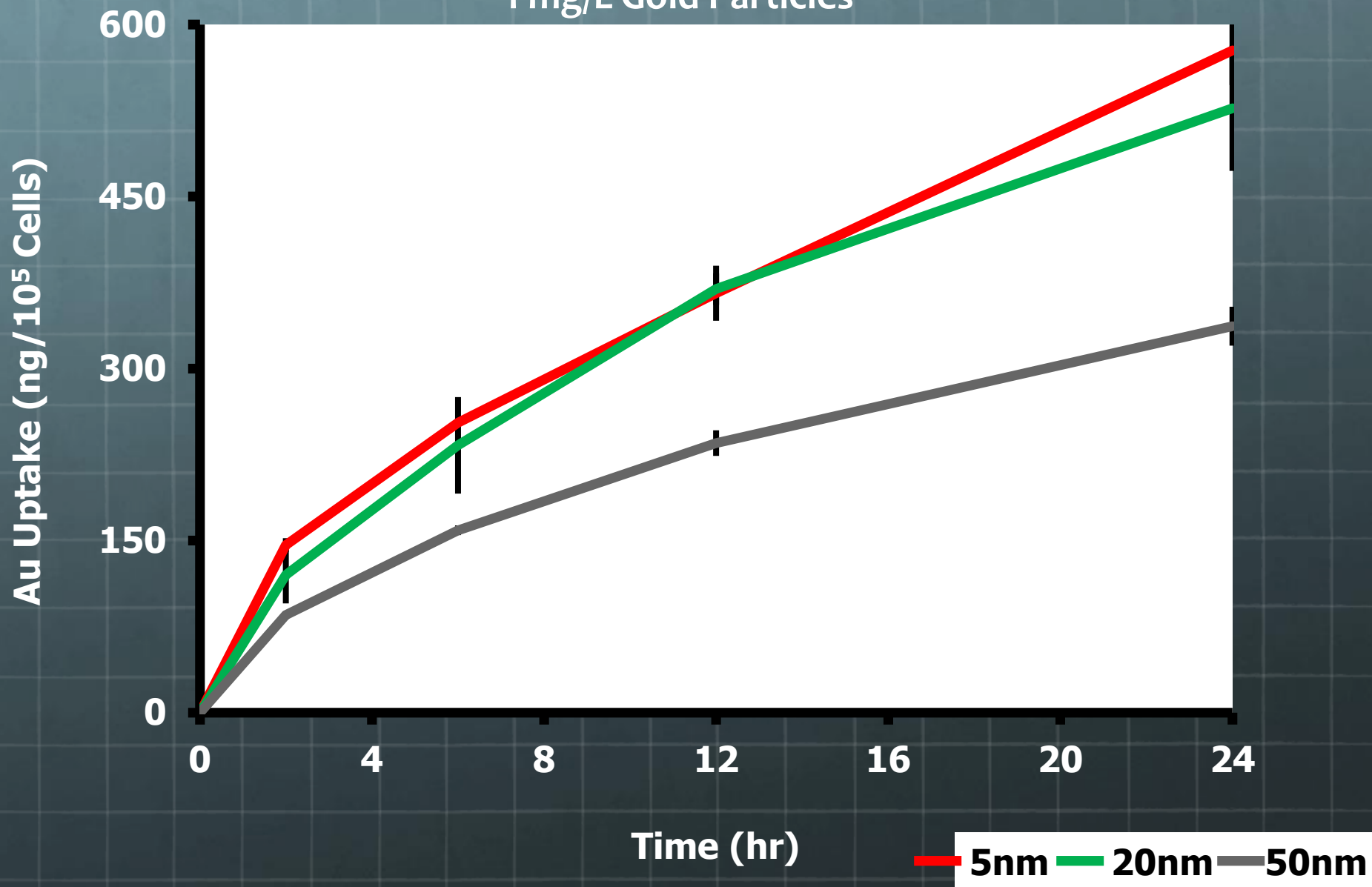
5% FBS

10% FBS

20% FBS

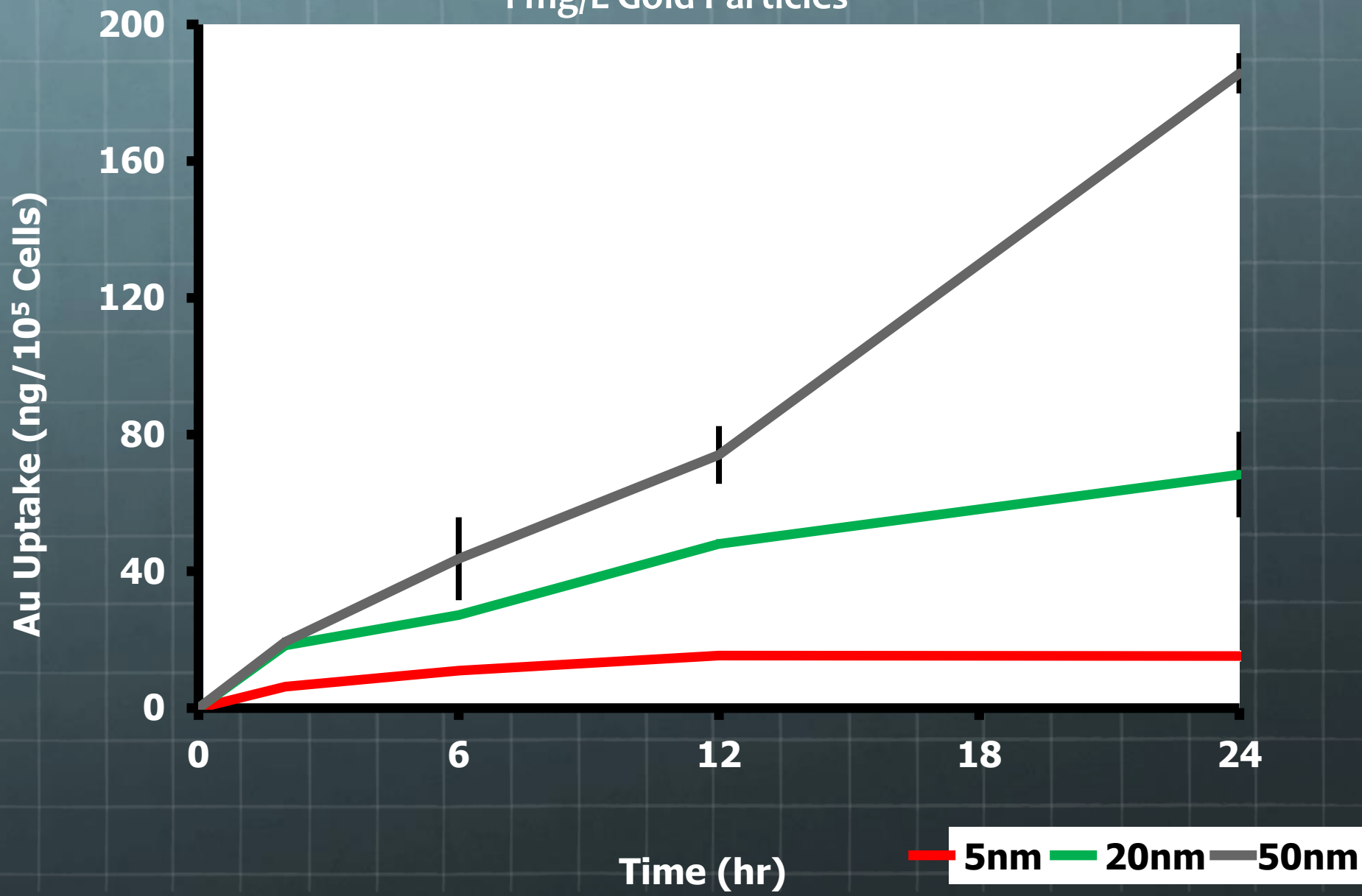
5% FBS

1 mg/L Gold Particles



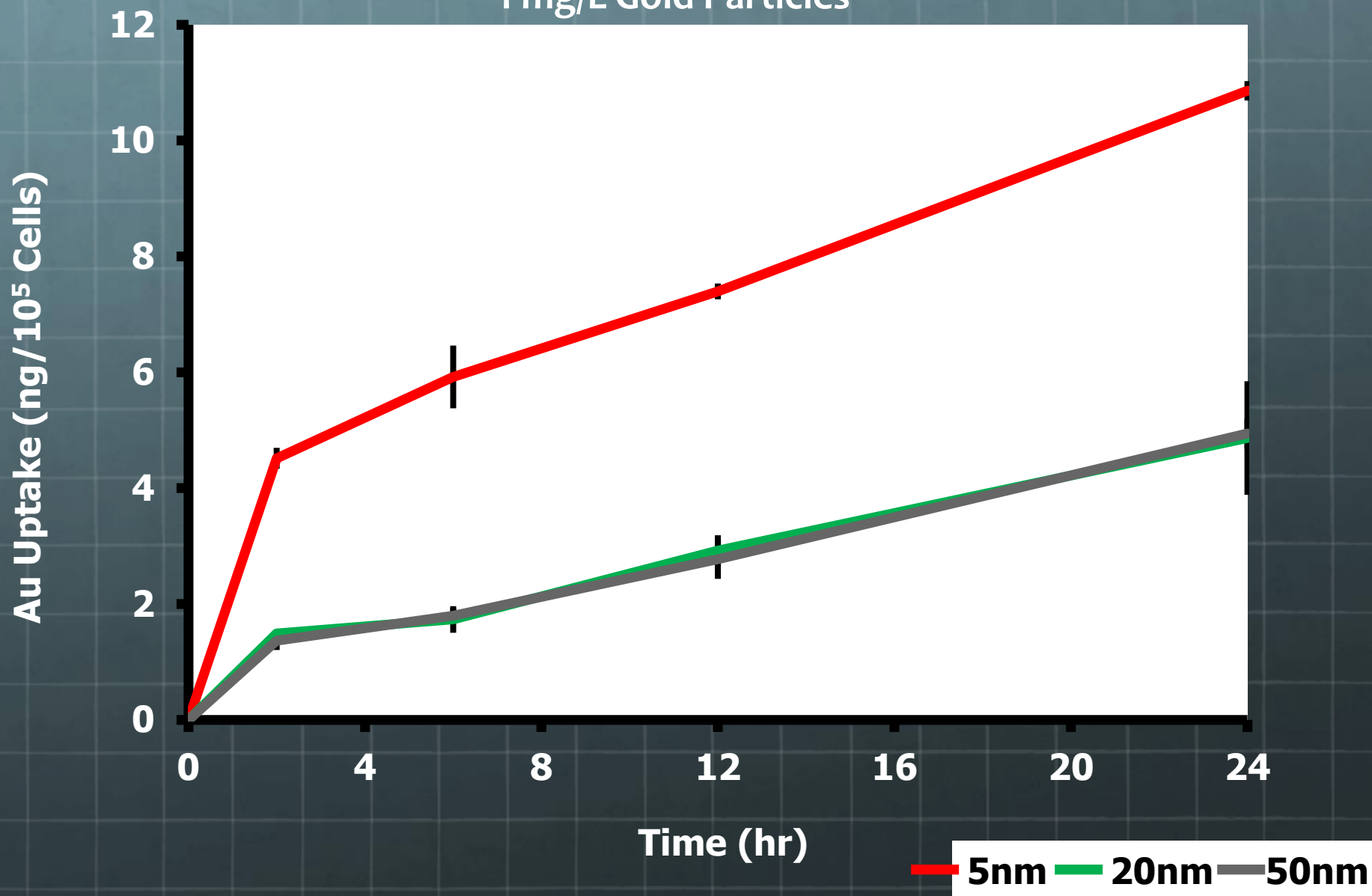
10% FBS

1 mg/L Gold Particles

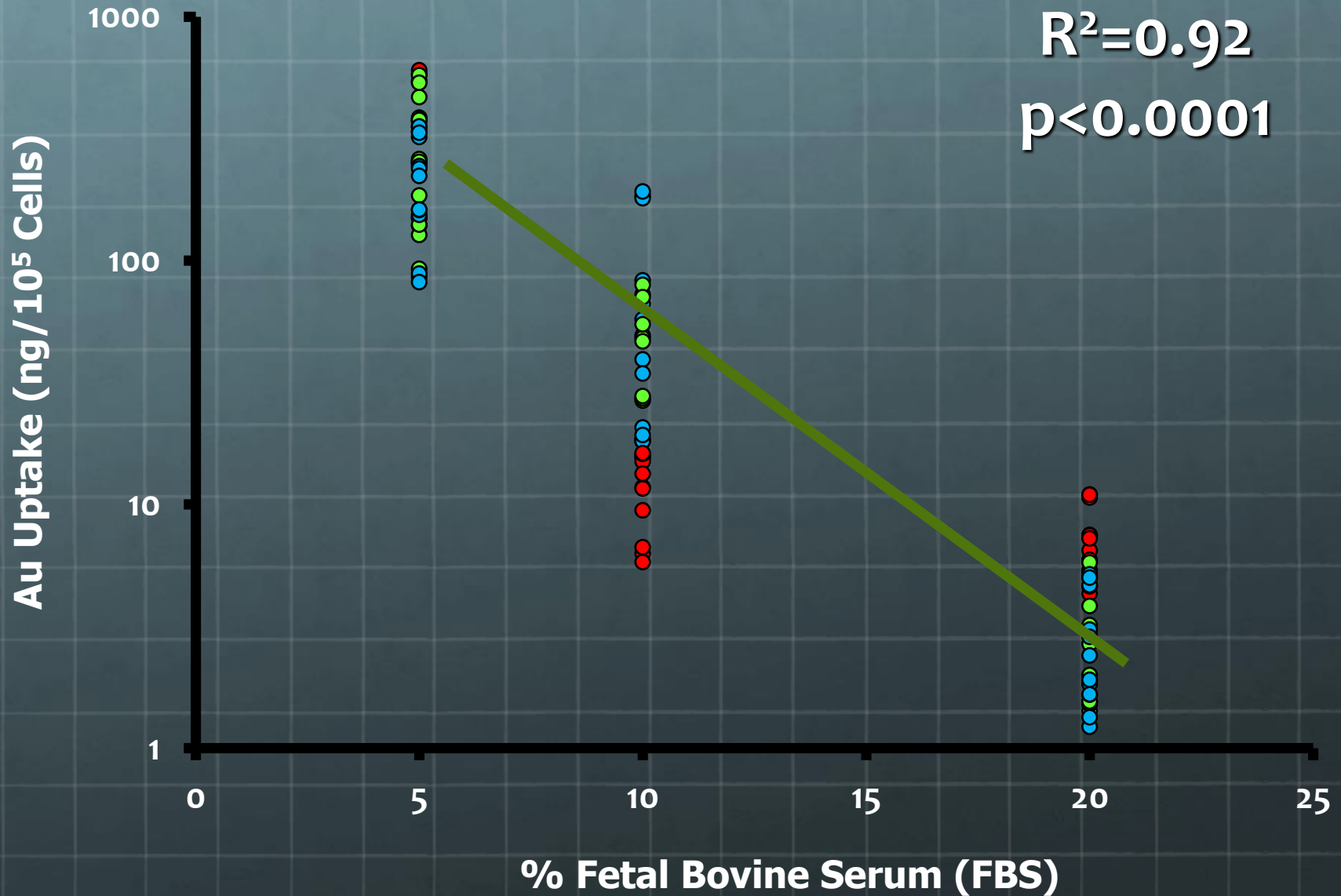


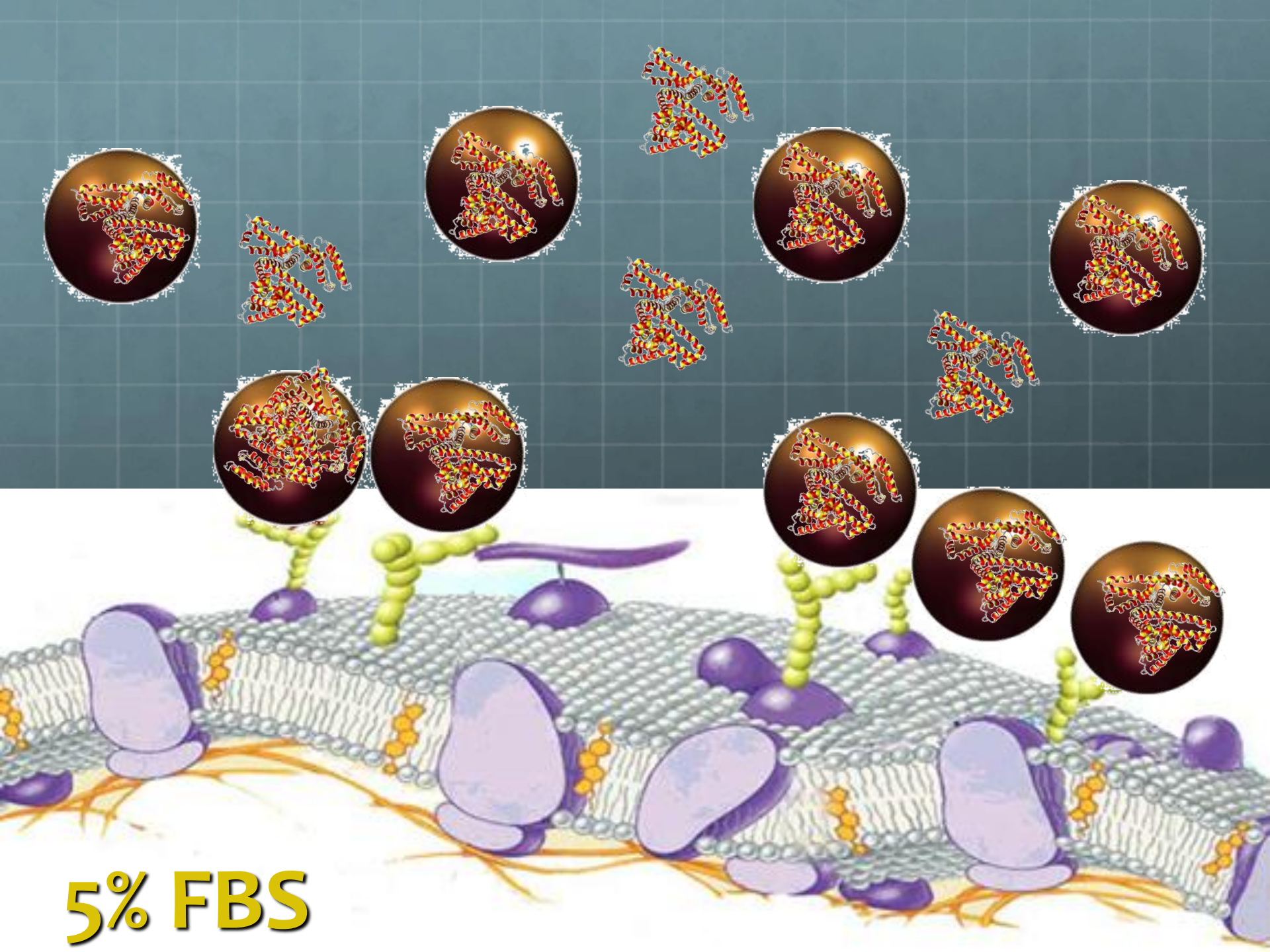
20% FBS

1 mg/L Gold Particles

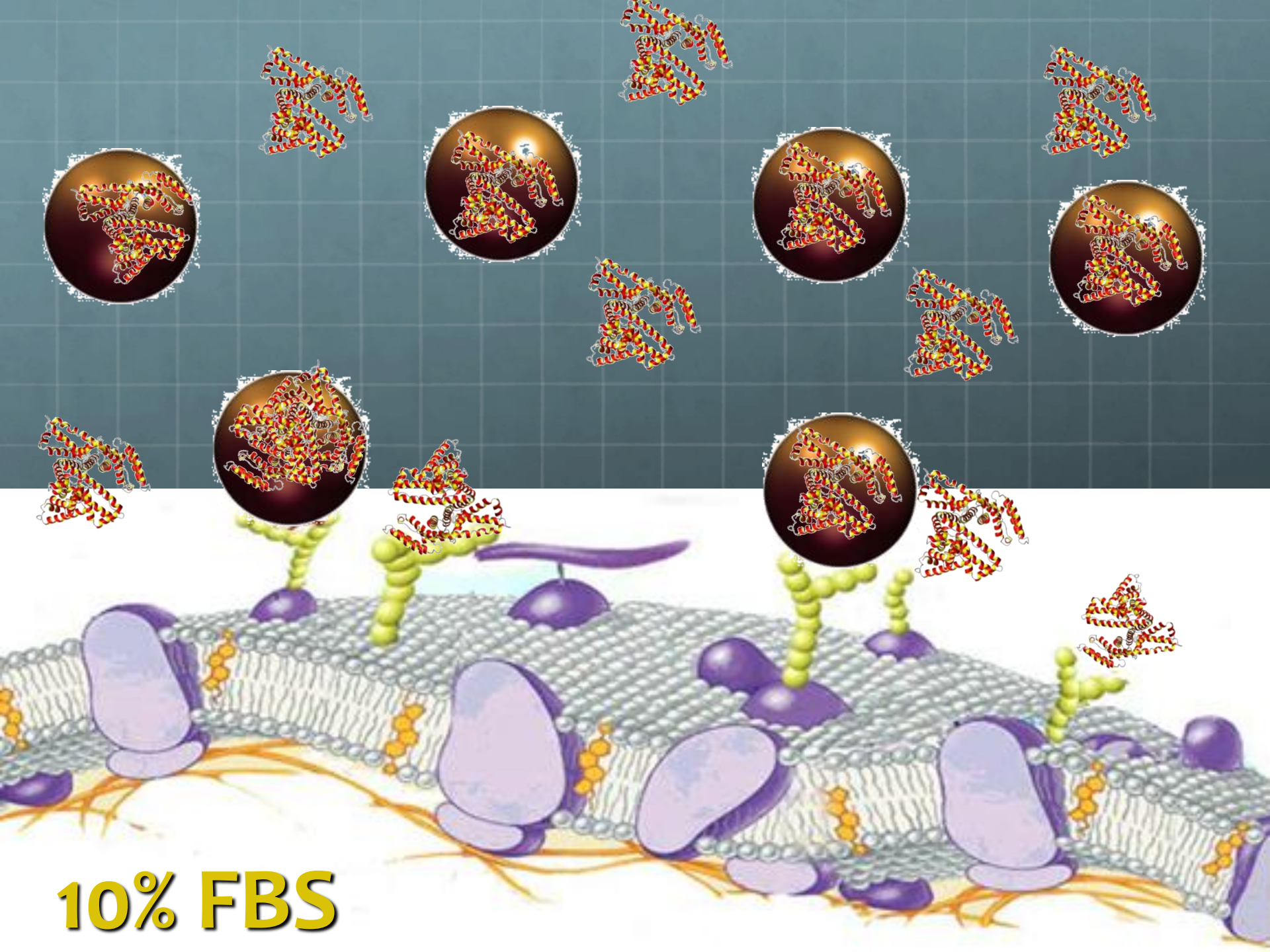


Regression

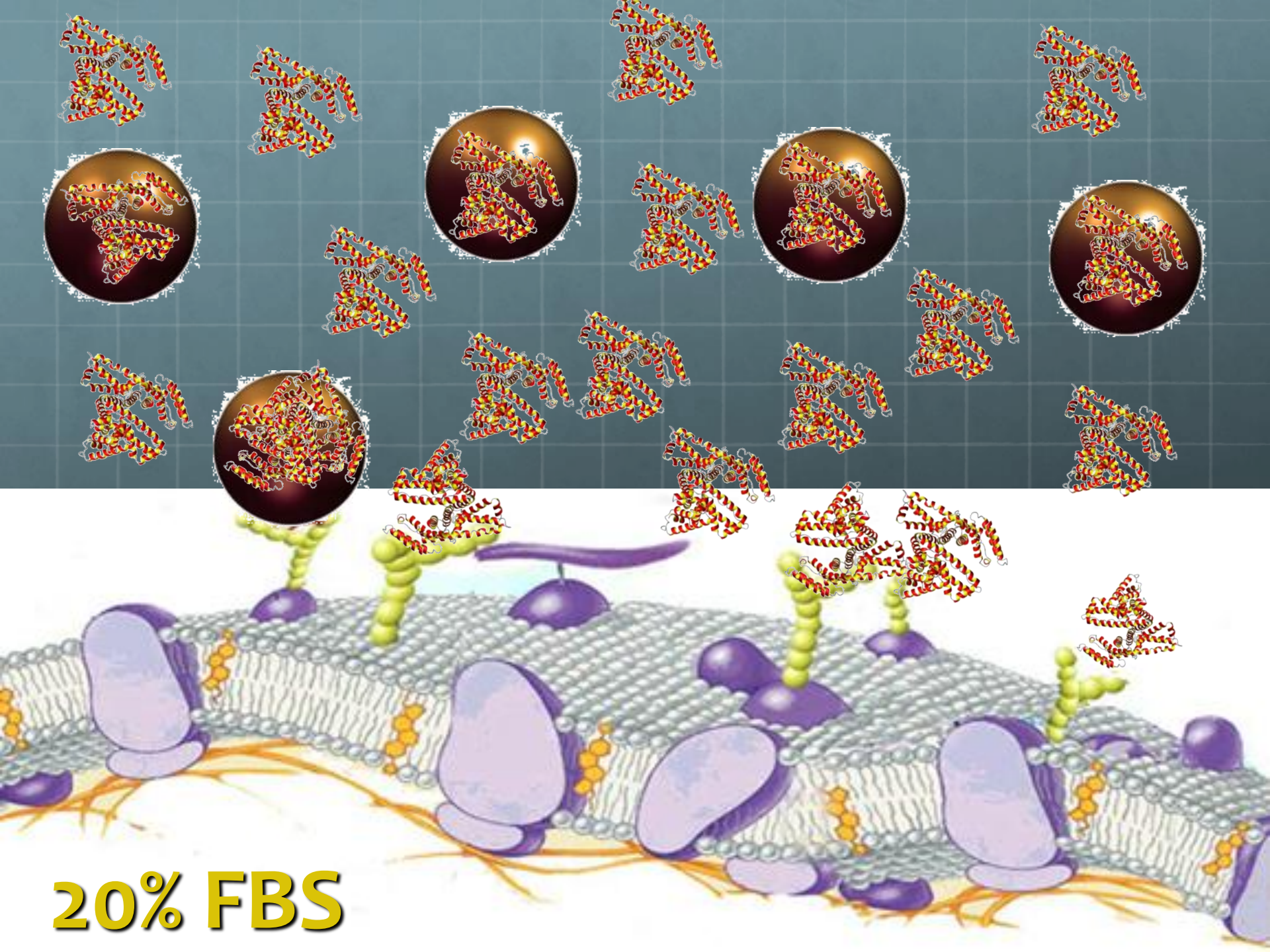




5% FBS



10% FBS



20% FBS